

SOME 19-NORGIBBERELL-16-ENES AS INHIBITORS OF GIBBERELLIN

PLANT HORMONE BIOSYNTHESIS

Mark K. Baynham and James R. Hanson \*

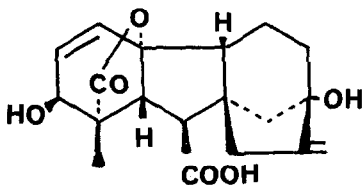
School of Molecular Sciences, University of Sussex, Brighton, Sussex, BN1 9QJ

**Abstract:** Methyl ent-7-hydroxy-4 $\beta$ (H)-19-norgibberell-16-en-20-oate, the corresponding 7,20-diol and 20-desoxy derivative, have been prepared from gibberellins A<sub>13</sub> and A<sub>14</sub> respectively and shown to act as inhibitors of gibberellic acid biosynthesis in Gibberella fujikuroi; further investigation of the 7-hydroxy-20-methyl ester suggests that this may be acting at the ring contraction step.

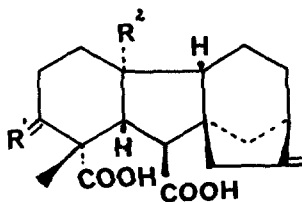
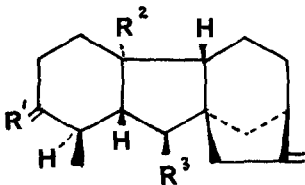
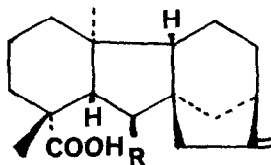
We have recently shown that ent-kauran-16,17-epoxide<sup>1</sup> and some B-norkaurenes<sup>2</sup> act as mimics of intermediates in gibberellin biosynthesis and, by blocking key steps in the formation of these plant hormones, lead to novel plant growth regulators. The C-19 carboxyl group plays an important role in several biosynthetic steps, providing for example, the lactone ring of the C<sub>19</sub> gibberellins such as gibberellic acid (1). We have prepared some 19-norgibberell-16-enes lacking this group and investigated them as potential inhibitors of the formation of gibberellic acid (1) in Gibberella fujikuroi.

Gibberellins A<sub>13</sub> (2)<sup>3</sup> and A<sub>14</sub> (3)<sup>4</sup> were oxidized to their  $\beta$ -ketones (4 and 5) and the resultant  $\beta$ -keto-acids, decarboxylated. The C-3 carbonyl group was then removed by Wolff-Kishner reduction and the  $\beta$ -desoxy acids methylated to afford (6) and (7). In the case of gibberellin A<sub>13</sub> a cleaner product was obtained by methylation of the 19-nor- $\beta$ -ketone (8), reduction to the  $\beta$ -alcohol (9) with sodium borohydride under carefully controlled conditions, conversion to the  $\beta$ -chloro compound with triphenylphosphine and carbon tetrachloride and reduction of (10) with tri-n-butyl tin hydride to afford the  $\beta$ -desoxy compound (6). Reduction of the 7- and 20-ester groups with lithium aluminium hydride proceeded in a stepwise manner in the gibberellin A<sub>13</sub> series to afford the

7-mono-ol (11) (major product) and the 20-mono-ol (12) (minor product) and the 7,20-diol (13). In the gibberellin A<sub>14</sub> series reduction of the ester (7) gave the 7-mono-ol (14). The stereochemistry of the 4-methyl group followed from the <sup>1</sup>H n.m.r. spectrum of (15) in which  $J_{4:5} = 12.5$  Hz and  $J_{5:6} = 12$  Hz (8 0.93 H-18; 2.80, H-4; 2.40, H-5; 3.52, H-6).



1

2 R<sup>1</sup> = α-H, β-OH; R<sup>2</sup> = CO<sub>2</sub>H3 R<sup>1</sup> = α-H, β-OH; R<sup>2</sup> = Me4 R<sup>1</sup> = :O; R<sup>2</sup> = CO<sub>2</sub>H5 R<sup>1</sup> = :O; R<sup>2</sup> = Me6 R<sup>1</sup> = H<sub>2</sub>; R<sup>2</sup> = R<sup>3</sup> = CO<sub>2</sub>Me7 R<sup>1</sup> = H<sub>2</sub>; R<sup>2</sup> = Me; R<sup>3</sup> = CO<sub>2</sub>Me8 R<sup>1</sup> = :O; R<sup>2</sup> = R<sup>3</sup> = CO<sub>2</sub>H9 R<sup>1</sup> = α-OH, β-H; R<sup>2</sup> = R<sup>3</sup> = CO<sub>2</sub>Me10 R<sup>1</sup> = β-Cl, α-H; R<sup>2</sup> = R<sup>3</sup> = CO<sub>2</sub>Me11 R<sup>1</sup> = H<sub>2</sub>; R<sup>2</sup> = CO<sub>2</sub>Me; R<sup>3</sup> = CH<sub>2</sub>OH12 R<sup>1</sup> = H<sub>2</sub>; R<sup>2</sup> = CH<sub>2</sub>OH; R<sup>3</sup> = CO<sub>2</sub>Me13 R<sup>1</sup> = H<sub>2</sub>; R<sup>2</sup> = R<sup>3</sup> = CH<sub>2</sub>OH14 R<sup>1</sup> = H<sub>2</sub>; R<sup>2</sup> = Me; R<sup>3</sup> = CH<sub>2</sub>OH15 R<sup>1</sup> = O; R<sup>2</sup> = R<sup>3</sup> = CO<sub>2</sub>Me16 R = CH<sub>2</sub>OH

17 R = CHO

Our strain of Gibberella fujikuroi (ACC 917) produces gibberellic acid (1) at a concentration of approximately 40 mg. litre<sup>-1</sup> and hence the 19-norgibberellenes were tested at this level. The hydroxy-ester (11), the diol (13) and the mono-ol (14) substantially (67 - 80%) inhibited the incorporation of [2-<sup>14</sup>C] mevalonic acid into gibberellic acid (1) (see table )

Table: % Incorporation of [2-<sup>14</sup>C]MVA into (1) and (2)

Substrate	Metabolite	
	Gibberellic acid (1)	Gibberellin A <sub>13</sub> (2)
Control	2.44	0.128
(11)	0.47	0.016
(13)	0.61	0.051
(14)	0.81	0.086

10 $\mu$ C 2-<sup>14</sup>C MVA and 40 mg substrate were incubated with G. fujikuroi in 1 litre medium (50 ml per flask) on shake culture for 4 days.

Gibberellin A<sub>13</sub> (2) is a terminal metabolite on the C<sub>20</sub> pathway. If these three compounds were blocking the loss of C-20 and the formation of the 19  $\rightarrow$  10 lactone, we would expect some additional radioactivity in the gibberellin A<sub>13</sub> in the presence of the inhibitors. Instead the amount of radioactivity in the gibberellin A<sub>13</sub> decreased and hence inhibition was prior to this step. The mono-hydroxy-ester (11) was therefore investigated further. Gibberellin A<sub>12</sub> 7-alcohol (16), although not a true intermediate in gibberellin biosynthesis, is a stable relative of the first gibbane intermediate, gibberellin A<sub>12</sub> 7-aldehyde (17). It is converted into gibberellic acid in comparable yield to the aldehyde.<sup>5</sup> The mono-hydroxy-ester (11) had no effect on the incorporation of [17-<sup>14</sup>C]-gibberellin A<sub>12</sub> alcohol (16) into gibberellic acid (1) ( 8.89% incorporation in the control, 8.71% incorporation in the presence of the substrate). In contrast the incorporation of ent-[<sup>14</sup>C]-kaurene was inhibited (0.18% in the control, 0.04% in the presence of the substrate). A radio-active metabolite with the t.l.c. characteristics of ent-7 $\alpha$ -hydroxykaurenoic acid accumulated. Consequently this 19-norgibberell-16-ene

is blocking gibberellin biosynthesis at a stage in the oxidative metabolism of ent-kaurene, possibly that of ring contraction. In this context it is interesting to note that although esters of ent-kaur-16-en-19-oic acid in which the 19-carboxylic acid is blocked, are hydroxylated on ring B they do not undergo ring contraction unless they are readily hydrolysed.<sup>6,7</sup> It is possible that the 19-carboxylic acid plays a more important role in this step than has hitherto been realized.

#### References

1. J.R.Hanson, C.L.Willis and K.P.Parry, Phytochemistry, 1980, 19, 2323
2. J.R.Hanson, K.P.Parry and C.L.Willis, J.C.S.Chem.Commun., 1981, 285; Phytochemistry, 1982, 21, 1575; idem, 1982, 21; 1955; J.R.Hanson, K.P.Parry, J.Triana and C.L.Willis, J.C.S.Chem.Commun., 1982, 192; Phytochemistry, 1983, 22, 97.
3. R.H.B.Galt, J.Chem.Soc., 1965, 3143.
4. B.E.Cross, J.Chem.Soc., 1966, 501.
5. J.R.Hanson and J.Hawker, Phytochemistry, 1973, 12, 1073.
6. P.R.Jefferies, J.R.Knox and T.Ratajczak, Phytochemistry, 1974, 13, 1423.
7. J.R.Hanson and F.Y.Sarah, unpublished work.

We thank I.C.I.Plant Protection PLC for a gift of materials and Drs D.Griffin and K.P.Parry for discussions.

(Received in UK 7 February 1983)